# TOTAL CONCENTRATION OF FREE AMINO ACIDS AND THE CORRESPONDING MORPHOGENETIC CHANGES DURING THE EMBRYOGENESIS OF CHILOMENESE SEXMACULATA FABR. (COLEOPTERA: COCCINELLIDAE) 

Devinder Mukesh, Dalbinder Singh Sidhu ${ }^{1}$ and Nirmal Kumar

Abstract.-The concentration of free amino acid (FAA) pool in relation to the various morphogenetic events occurring during embryogenesis of Chilomenes sexmaculata Fabr. was determined at different incubation periods. During the first 12 hours of embryogenesis, when the zygote undergoes an active cleavage, the concentration declines gradually. However, it rises sharply at 15 hours of development when the cleavage energids undergo an active migratory activity. The amount of FAA decreases at 24 hours and remains almost steady up to 35 hours of incubation when the embryo starts elongation and segmentation. From then on there is a continuous increase in FAA up to 75 hours followed by a declining trend towards hatching as the yolk gets depleted.

## Introduction

Extensive reviews on the biochemical investigations pertaining to the nitrogen metabolism during embryogenesis of insects have earlier been made by Chen $(1962,1966)$ and Corrigan (1970). To get a greater insight into the development of an embryo it is necessary to elucidate the relationship of biochemical changes to the corresponding morphogenetic events occurring during this period. Since the major processes underlying differentiation at the embryonic level aim at protein synthesis which in turn depends on the supply of free amino acids (FAA) from the yolk reserves, FAA thus have a distinct relationship to morphogenesis.

The present attempt unveils a relationship between the total concentration of FAA and the corresponding morphogenetic events occurring during the different incubation periods in Chilomenes sexmaculata Fabr.

## Materials and Methods

The adults of $C$. sexmaculata were collected from the Calotrips procera plants during the months of November to February around Patiala (India) and were reared in the laboratory according to Sarswat (1976) on Aphis neri. Numerous batches of eggs were obtained, incubated at $28 \pm 2^{\circ} \mathrm{C}$ for

[^0]Table 1. Concentration of FAA pool and various morphogenetic events occurring during embryogenesis of Chilomenes sexmaculata Fabr.

| Incubation period (hr) | $\mu \mathrm{M}$ FAA/ 100 eggs $( \pm$ SD)* | Morphogenetic event |
| :---: | :---: | :---: |
| 0.25 | $1.51 \pm 0.030$ | Maturation of the male as well as female pronucleus (Fig. 1). |
| 1.00 | $1.48 \pm 0.029$ | Fusion of male and female nucleus (Fig. 2). |
| 3.00 | $1.34 \pm 0.027$ | Cleavage of the zygote, giving rise to 4 -energids stage (Fig. 3). |
| 5.00 | $1.46 \pm 0.029$ | 8-cleavage energids stage (Fig. 4). |
| 7.50 | $1.16 \pm 0.023$ | The process of cleavage continues. |
| 12.00 | $1.11 \pm 0.022$ | $240 \pm 16$ energids stage (Fig. 5). |
| 15.00 | $1.77 \pm 0.035$ | Migration of the cleavage energids towards the periplasm starts (Fig. 6). |
| 19.00 | $1.68 \pm 0.035$ | A syncytial primary epithelium is formed (Fig. 7). |
| 24.00 | $1.37 \pm 0.028$ | A regular layer of primary epithelial cells observed (Fig. 8). |
| 31.00 | $1.39 \pm 0.028$ | The ventro-posterior portion of the primary epithelium becomes thickened, the cells being columnar, so as to give rise to the definitive germ primordium (ventral plate) (Fig. 9). |
| 35.00 | $1.32 \pm 0.026$ | Elongation of the germ band or ventral plate takes place and the germ band becoming faintly distinguishable into an anterior protocephalon and the posterior protocorm (Fig. 10). |
| 45.00 | $1.42 \pm 0.028$ | Further elongation of the germ band resulting in the formation of three protocormic regions (Fig. 11). |
| 60.00 | $1.45 \pm 0.029$ | The germ band becomes segmented with delineated appendages (Fig. 12). |
| 72.00 | $1.48 \pm 0.029$ |  |
| 75.00 | $1.55 \pm 0.030$ | An active growth period, the appendages further develop and the process of cephalization starts, secondary yolk cleavage, yolk poly-hedra formed. |
| 90.00 | $1.35 \pm 0.026$ | The appendages become fully formed and the head also becomes distinct with the usual appendages (Fig. 13). |
| 100.00 | $1.30 \pm 0.026$ | Fully formed embryo. |
| 110.00 | $1.22 \pm 0.024$ | Hatching takes place. |

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Fig. 1. Abbreviations: CN, cleavage energid; FN, female pronucleus; G, germ band; GA, gnathal appendages; LR, labral appendages; MN, male pronucleus; PC, pole cell; PE, primary epithelium; PM, protocorm; 1PM, 2PM, 3PM, 1st, 2nd and 3rd protocormic regions; PMB, premandibular appendages; PN, protocephalon; TA, thoracic appendages; YC, yolk cell.
definite periods and subsequently fixed in Carl's fixative at $60^{\circ} \mathrm{C}$ for 2 hours. The permanent preparations were made according to the method of Rempel and Church (1971) to observe various developmental changes in the embryo.

The FAA extracts from the various egg samples at specific incubation periods were prepared according to Sidhu and Kang (1979). The quantitation of FAA pool was made by the method of Troll and Cannon (1953).

## Results and Discussion

The concentration of FAA pool and the various morphogenetic events occurring during the embryogenesis of Chilomenes sexmaculata Fabr. are recorded in Table 1.

At 0 hour, i.e., immediately after oviposition, the concentration was $1.51 \pm 0.030 \mu \mathrm{moles} / 100$ eggs, which decreased three hours later to $1.34 \pm$ $0.027 \mu$ moles $/ 100$ eggs. This decrease in FAA concentration corresponds to the period when the exochorion of the newly laid egg becomes hard and
dark in color, apparently indicating the utilization of FAA in this process. It has further been noted that there is a continuous decrease in FAA concentration up to 12 hours of incubation, i.e., during the active cleavage activity when the zygote nucleus undergoes multiple synchronous divisions. This decrease suggests that the breakdown of yolk proteins has not yet started and the need of amino acids for protein synthesis is met from the already existing FAA pool accumulated during maturation of egg. The initiation of protein synthesis immediately after fertilization has earlier been reported by Lockshin (1966) in some coleopterous insects.

At about 15 hours of incubation, when energids start migrating, the concentration of FAA suddenly increases to a fairly high value ( $1.77 \pm 0.035$ $\mu$ moles $/ 100 \mathrm{egg}$ ). This increase is due perhaps to the lysis of yolk proteins, with the result that the mixing-up of the yolk contents facilitates the peripheral migration of the cleavage energids. This type of increase in FAA concentration has been observed by Chen and Briegel (1965) in Culex pipens morestus at a stage when blastoderm formation and elongation of germ band (Idris 1960) takes place.

The FAA concentration sharply decreases ( $1.37 \pm 0.28 \mu \mathrm{moles} / 100$ eggs) at 24 hours of incubation when a regular primary epithelial layer gets established, revealing a higher rate of protein synthesis in comparison with the breakdown of yolk proteins. Thereafter the concentration remains almost steady up to 34 hours of development. During this period the differentiation of the ventral plate or the germ band is completed.

At 45 hours of development the amount of FAA again rises ( $1.42 \pm 0.028$ $\mu$ moles $/ 100$ eggs), when the germ band undergoes further elongation and primary segmentation. This trend in FAA concentration increase was observed in the following 30 hours of development. The peak is reached at 75 hours of incubation when the embryo shows an active development. This increase in concentration can be attributed to an active lysis of the yolk proteins, corresponding to an active growth period when cephalization as well as the phenomenon of secondary yolk cleavage takes place.

After this period, the FAA concentration shows a continuous decline when the appendages become fully formed and the head becomes quite distinct. This decline in concentration prior to hatching is due perhaps to the fact that the yolk reserves become increasingly exhausted at this stage and the supply of amino acids can no longer keep pace with the demand for more amino acids for protein synthesis. Chen (1966) and Indira (1963) have also observed a similar type of decrease in FAA concentration towards hatching.

## Literature Cited

Chen, P. S. 1962. Free amino acids in insects; in Holden: Amino Acids Pools, pp. 115-138. Elsevier, Amsterdam.
—_. 1966. Amino acid and protein metabolism in insect development. Adv. Insect Physiol. 3:53-132.
___ and H. Briegel. 1965. Studies on protein metabolism of Culex pipiens L. V. Changes in free amino acids and peptides during embryonic development. Comp. Biochem. Physiol. 14:463-473.
Corrigan, J. J. 1970. Nitrogen metabolism in insects; in Campbell: Comparative biochemistry of nitrogen metabolism. I. The invertebrates, pp. 387-488. Academic Press, London.
Indira, T. 1963. Biochemical and cytological studies during development and ovarian growth in Sphaeroderma malestum (Duf.). Ph.D. Thesis, Annamalai University, South India.
Idris, B. E. M. 1960. Die Entwicklung im novmalen Ei von Culex pipiens (Diptera). Z. Morph. Ökol. Tiere 49:387-429.
Lockshin, R. A. 1966. Insect embryogenesis: Macromolecular synthesis during early development. Science 154:775-776.
Rempel, J. G. and N. S. Church. 1971. Embryology of Lytta viridana LeConte (Coleoptera: Meloidae). III Appendiculate 72h embryo. Can. J. Zool. 49:1563-1571.
Sarswat, G. G. 1976. Studies on the trends in consolidation of nervous system in Coleoptera. Ph.D. thesis, Agra University, Agra (India).
Sidhu, D. S. and H. K. Kang. 1979. Metabolic reserves and pool size of free amino acids during metamorphosis of Callosobruchus maculatus (F.). Entomon. 4(1):57-59.
Troll, W. and R. K. Cannon. 1953. A modified photometric ninhydrin method for the analysis of amino and imino acids. J. Biol. Chem. 200:803.

Department of Zoology, Punjabi University, Patiala-147002, India.
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[^0]:    ${ }^{1}$ All correspondence to this author.

[^1]:    * Standard deviation. Each reading is an average of three determinations.

